



# ZOONO TECHNICAL OVERVIEW

## CHEMICAL STRUCTURE & NOMENCLATURE

### Compound (ODTA)

Octadecyldimethyl (3-trimethoxysilylpropyl) ammonium chloride

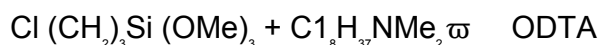
### Structure



**Note:** This material is classified as an organofunctional trihydroxysilane as it contains a functional organic group (quaternary nitrogen) covalently bound to a silicon atom. Organosilane denotes a minimum of one carbon-silicon bond. Trihydroxy describes the number of hydroxy groups bound to the silicon atom.

### General Silane Chemistry

ODTA Prep.

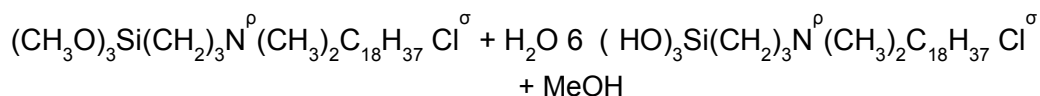


## ZOONO CHEMISTRY

Zoono is prepared as a trialkoxysilane. Trialkoxysilanes undergo a wide variety of chemical reactions including trans-esterification, acetylation, halogenation, condensation and hydrolysis to name but a few. The most important of these reactions are hydrolysis (which is used to prepare Zoono) and condensation, (which is required for Zoono to bond to surfaces and form polymeric films).

### Hydrolysis of ODTA to Zoono

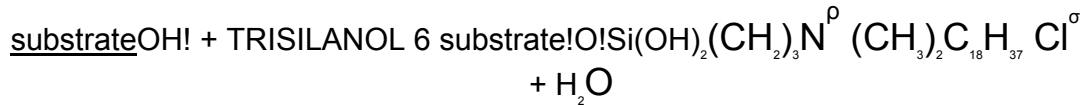
- A. On contact with water molecules, the methoxy groups attached to the silicon atom in the ODTA molecule will react in a step-wise manner to form silanols.



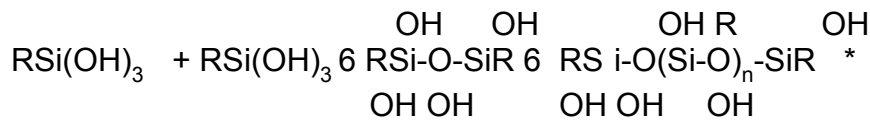
(In dilute aqueous solution, a stable solution is formed).

## ZOONO CHEMISTRY – Contd...

- B. The Silanol moieties are very reactive and will react through a condensation type reaction with a wide variety of hydroxylated surfaces including cellulose, metals, sand, silica, zeolites, etc.

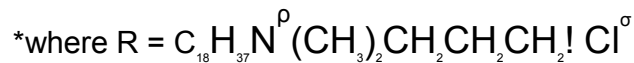
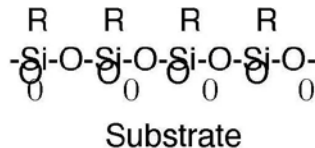


- C. Silanol containing Zoono undergo autocondensation reactions with other Silanol substituted Zoono molecules and with the surface of a substrate, eliminating water.



Autocondensation occurs across the surface of substrates to produce 3-dimensional, cross-linked polymers being on the average 10-30 molecules thick, depending on the substrate.

Drying after application produces a durable, non-leaching, bonded, antimicrobial surface.



## GENERAL REACTIVITY OF ZOONO

### Bonding and Deposition on Surfaces

Zoono will react with oxide and hydroxyl containing surfaces through a condensation reaction. The bonds formed are covalent and are resistant to re-hydration. Coatings on siliceous materials (glass, sand) are removed by thermal means (>250°C) or concentrated chemical treatment (KOH, NaOH, HF). Durable coatings on metal are dependent on the metal substrate.

Zoono coatings have been applied to textile fibres including cotton, cellulose acetate, polyester, nylon, wool, rayon, acrylon, etc.

**Bonding and Deposition on Surfaces – Contd...**

Durable coatings are readily applied to various natural and plastic surfaces including cotton, cellulose, wool, nylon, PVC, polyester, rayon, etc. Evaporation of the application media promotes T-resin film formation on the substrate.

Electrostatic or H-bonding interactions provide bonding. Uniform film formation occurs through even “wetting” of the plastic surface. Durable films are achieved on nylon, fibreglass, epoxy, polyester, PVC substrates. Less durable films are obtained on PE and PP. Film formation on PTFE and other fluorinated substrates have not been reported.

**GENERAL ZOONO ANTIMICROBIAL CHEMISTRY**

Zoono is a unique antimicrobial material.

It forms a water stable solution at low concentrations. There are no additives in Zoono added to promote stability. This solution has the antimicrobial properties of typical aqueous solutions of quaternary ammonium compounds.

On application to surfaces, the material reacts to produce a bonded antimicrobial coating. On evaporation of the water solvent, a durable bonded coating is produced which has improved and sustained antimicrobial properties.

Zoono, when applied to a surface, transforms from a water soluble monomeric species to an insoluble, macromolecular antimicrobial film.

This film is stable and durable.

Zoono has been demonstrated effective against a wide range of pathogens.

## APPENDIX ONE

The following table lists the pathogens that are killed or inactivated following application of Zoono to various substrates and materials.

This table does not reflect the activity of Zoono in the liquid form.

### Pathogens Inactivated By Zoono Application

<b>Gram Positive Bacteria</b>	<b>Reference</b>
Bacillus sp. (vegetative cell)	5, 6, 11
Corynebacterium diphtheriae	1, 13
Micrococcus lutea	5, 6, 11
Micrococcus s p.	2, 5, 15
Mycobacterium tuberculosis	14, 36
Mycobacterium smegmatis	14
Propionibacterium acnes	5
Staphylococcus aureus *	2, 3, 5, 6, 10, 11, 13, 24, 15, 21
Staphylococcus epidermidis	2, 5, 6, 7, 11, 13, 14, 15
Streptococcus faecalis	2, 5, 6, 7, 11, 13, 14
Streptococcus mutans	5, 6, 7, 11
Streptococcus pneumonia	1
Streptococcus pyogenes	5, 6, 7, 11
<b>Gram Negative Bacteria</b>	<b>Reference</b>
Acinetobacter calcoaceticus	2, 5, 6, 11, 14, 15
Aeromonas hydrophilia	5, 6, 11
Citrobacter deversus	5, 6, 11
Citrobacter freundii	5, 6, 11
Enterobacter aerogenes	5, 6, 7, 11
Enterobacter agglomerans	2, 5, 14, 15
Enterobacter cloacae	5, 6, 7, 11
Enterococcus	10
Escherichia coli	1, 2, 3, 5, 6, 7, 10, 11, 13, 14
Klebsiella oxytoca	5, 6, 11, 14
Klebsiella pneumoniae	3, 5, 6, 7, 9, 10, 11, 13, 14
Klebsiella terrigena	19
Legionella pneumophila	1
Morganella morganii	5, 6, 7, 11
Proteus mirabilis	5, 6, 7, 11
Proteus vulgaris	5, 6, 7, 11
Pseudomonas aeruginosa	2, 3, 5, 6, 7, 11, 13, 14
Pseudomonas fluorescens	5, 6, 7, 10, 11
Salmonella cholerae suis	5, 6, 7, 11, 14
Salmonella typhi	5, 6, 7, 11, 14
Salmonella typhimurium	1, 5, 6, 7, 11
Serratia liquifaciens	5, 6, 7, 11
Serratia marcescens	5, 6, 7, 11
Xanthomonas campestris	5, 6, 7, 11

Contd...

## **Viruses**

## **Reference**

Adenovirus Type II & IV	17, 18, 21
Bovine Adenovirus Type I & IV	17, 18, 21
Feline pneumonitis	21
Herpes Simplex Type I	16, 17, 18
Herpes Simplex Type II	21
HIV-1 (AIDS)	21
Influenza A2 (Aichi)	17, 18, 21
Influenza A2 (Asian)	17, 18
Influenza B	17, 18
Mumps	17, 18
Parinfluenza (Sendai)	21
Rous S arcoma	17, 18
Reovirus Type I	17, 18
Simian Virus 40	17, 18
Vaccinia	17, 18
MS2	19
PRD1	19
H1N1 (Swine Flu)	34
Norovirus	35
Middle East Respiratory Syndrome (MERS)	37
Ebola ** (See Note)	38

## **Fungi, Algae, Mould, Yeast, Spores**

## **Reference**

Alterania alternate	8,12
Aphanizomenon s p.	22
Aspergillus flavus	2, 5, 6, 7, 11, 14
Aspergillus niger	2, 5, 6, 7, 8, 11, 12, 13, 14
Aspergillus sydowi	5, 6, 7, 11
Aspergillus terreus	5, 6, 7, 11, 14
Aspergillus versicolor	2, 5, 6, 7, 11
Aspergillus verrucaria	14
Aureobasidium pullans	5, 6, 7, 8, 11, 12
Candida albicans	1, 2, 5, 6, 7, 14
Candida pseudotropocalis	5, 6, 7, 11
Chaetomium globsum	1
Cladosporium cladosporioides	8, 12
Chlorella vulgaris	19
Dreschlera australiensis	8, 12
Epidermophyton s p.	9
Gliomastix cerealis	8, 12
Gloeophyllum trabeum	5, 6, 7, 11
Microsporum s p.	9
Microsporum audouinii	21
Monilia grisea	8, 12
Oscillatoria	20
Penicillium chrysogenum	5, 6, 7, 11
Pencillium commune	8, 12
Penicillium funiculosum	1, 2, 5, 6, 7, 11, 14
Penicillium pinophilium	5, 6, 7, 11

Contd...

**Fungi, Algae, Mould, Yeast, Spores**

**Reference**

Penicillium variable	5, 6, 7, 11, 14
Phoma fimeti	8, 12
Pithomyces chartarum	8, 12
Poria placenta	5, 6, 7, 11
Scenedesmus	20
Saccharomyces cerevisiae	5, 6, 7, 11, 13, 14
Scolecobasidium humicola	8, 12
Selenastrum s p.	22
Trichoderma viride	5, 6, 7, 11
Trichophyton interdigitale	2, 14
Trichophyton maidson	14
Trichophyton mentogrophytes	5, 6, 7, 9, 11
Trichophyton sp.	9

**Protozoa Parasites**

**Reference**

Cryptosporidium parvum (oocysts)	19
----------------------------------	----

Note:

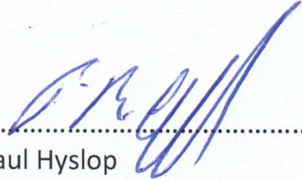
\*\* Special formulation containing H<sub>2</sub>O<sub>2</sub> + Zoono quaternary ammonium compound active

## REFERENCES

1. Y. Hsiao, Chinese Pat. Appl., PCT/CN98/00207 (1998)
2. James Malek, John Speir, "Method of Reducing the Number of Microorganisms in a Method of Preservation"; U.S. Pat. 4,259,103 (1981)
3. Stewart Klein, "3-(trimethoxysilyl)propyl-dodecylmethyl Ammonium Salts and Method of Inhibiting growth of Microorganisms Therewith"; U.S. Pat. 4,394,378 (1983).
4. William Eudy, "Organosilicon Quaternary Ammonium Antimicrobial Compounds"; U.S. Pat. 4,406,892 (1983).
5. Richard Gettings, William White, "Skin Treatment Method"; U.S. Pat. 4,908,355 (1990)
6. Lynne Blank, William White, "Antimicrobial Rinse Cycle Additive"; U.S. Pat. 5,145,596 (1992)
7. Richard Gettings, William White, "Ophthalmic fluid Dispensing Method"; U.S. Pat. 5,013,459 (1991).
8. Richard Avery, Frederick Martin, Sean Dwyer, "Production of Stable Hydrolyzable Organosilane Solutions"; U.S. Pat. 5,411,585 (1995).
9. Lynne Blank, Richard Gettings, William White, "Method of Treating Tinea Pedis and Related Dermatophytic Infections"; U.S. Pat. 4,865,844 (1989).
10. David Battice, Michael Hale, "Antimicrobially Effective Organic Foams and Methods for their Preparation"; U.S. Pat. 4,631,297 (1986).
11. Bruce Higgs, William White, "Solid Antimicrobial"; U.S. Pat. 5,359,104 (1994). This patent also describes the method of antimicrobial activity.
12. Richard Avery, Frederick Martin, Sean Dwyer, Colin Brown, "Production of Stable Hydrolyzable Organosilane Solutions"; U.S. Pat. 5,411,585 (1995).
13. William White, Jerry Olderman, "Antimicrobial Techniques for Medical Nonwovens: A Case Study"; Book of Papers, 1984, 12<sup>th</sup> Annual Nonwovens Tech. Symposium, pp. 13-46. No bacterial adaption (no increased bacterial resistance to Zoono) reported.
14. J. McGee, J. Malek, W. White, "New Antimicrobial Treatment for Carpet Applications", Am. Dyestuff Rep., 1983, (6), pp.56-59. Dow Corning Technical Brochure; 22-994-83 (1983).
15. Richard Gettings, Benny Triplett, "A New Durable Antimicrobial Finish for Textiles"; Book of Papers, 1978, American Association of Textile Chemists and Colorists National Technical Conference, pp. 259-261. Dow Corning Technical Brochure; 24-095-85 (1985).
16. I-Fu Tsao, Henry Wang, Charles Shipman, "Interaction of Infectious Viral Particles with a Quaternary Ammonium Chloride Surface"; Biotechnol. Bioeng, 34, (5), pp. 639-46 (1989).
17. I-Fu Tsao, Henry Wang, "Removal and Inactivation of Viruses by a Surface Bonded Quaternary Ammonium Chloride", ACS Symp. Ser. 1990, Volume Date 1988, 419, pp. 250-67. Reaction with Lipophilic Viruses.

## REFERENCES – Contd...

18. M. Klein, A. DeForest, "Principles of Viral Inactivation", Disinfection, Sterilization and Preservation. 3<sup>rd</sup> Ed., S. Block, Ed., (Lea & Febiger, Philadelphia, PA) 1983, pp.422-434.
19. M. Abbaszadegan, et. al., "Evaluation of Proprietary Treated Zeolite in Point of Use Devices for Removal of Microorganism", NSF Water Quality Center, Arizona State University, Tempe, AZ 85257; 12/03. W. Peterson & R. Berman, U.S. Pat.Pending, 60/472,429 (7/2003).
20. P. Westerhoff, D. Bruce, "Biocide Coating Experiment", Arizona State University, Tempe, AZ 85257; (2000).
21. W. Peterson, D. Giaccio, R. Berman, "Antimicrobial Skin Preparations Containing Organosilane Quaternaries", U. S. Patent 6,613,755 (9/2/03).
22. Third Party Testing; Univ. Iowa, Hygienic Laboratory, (No.27, AIHA, NELAD, USEPA, NVLAP), Iowa City, IA, (2005)
23. M. Abbaszadegan, et. al., J. Envir. Science & Health, Part A, 41:12011210, 2006.
24. A.J. Isquith, et.al., Applied Microbiology, 24, 859-863 (1972).
25. J.B. McGee, et .al., Am. Dyestuff Rep., 6: 56-59 (1983).
26. R.L. Gettings, AATCC, Book of Papers, 1978, p. 259
27. R.A. Kemper, W.C. White, R.L. Gettings, Dev. Indust. MicroBio., 31, (J.Ind. Micro Bio. 3,) p.237-244, (1990).
28. R.L. Getings, B.L. Triplett, AATCC Book of Papers, Natl. Tech. Conf. 1978. p. 259-261.
29. Third Party Testing; ZeroRez Franchising, Data Chem Labs., Salt Lake City,
30. UT, (2005).
31. Third Party Testing; SGS U.S. Testing Company Inc., Tulsa, OK(2004).
32. U.S. EPA, Press Release, (5/2001)
33. L.W. Dalton, Chemical & Engineering News, 2004, (2/16). 57-61.
34. MicroLab GmbH (Germany) Testing EN 14476:2007-02 (1). January 2010.
35. MikroLab GmbH (Germany) Testing (July 2010).
36. Abbott Analytical (UK) Testing May 2014 EN14348:2005
37. Institute for Hygiene & Microbiology (GmbH) June 2014 EN14476:2013
38. US Centre for Disease Control & Prevention 2014

  
.....  
Paul Hyslop  
CEO / MANAGING DIRECTOR,  
ZOONO GROUP LIMITED,  
AUCKLAND, NEW ZEALAND.

